Synthesis of 7,13-Bridged Arachidonic Acid Analogues

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cis- and trans-7,13-Bridged arachidonic acids (2 and 3, R=H) and their 5,6,14,15-tetradehydro-analogues (4 and 5, R=H) have been synthesized employing a divinylcyclopropane rearrangement as a key step.

As part of a programme directed towards modulating the arachidonic acid (1) system¹ we designed the bridged compounds (2)—(5) aiming to mimic the suspected bent J or U shape conformation of arachidonic acid² and diminish the reactivity of positions 7 and 10. We report here a facile and efficient entry into this class of compounds that involves a novel divinylcyclopropane rearrangement to construct the cycloheptadiene system.

Reduction of the *trans*-diester (6) (LiAlH₄, Et₂O, 0 °C, 90%) yielded the diol (7) which was converted by Swern oxidation [Me₂SO, (COCl)₂, Et₃N, CH₂Cl₂, 90%] into the *trans*-dialdehyde (8).³ Condensation of (8) with excess of the ylide derived from Me₃SiC \equiv CCH₂PPh₃Br⁴ (BuⁿLi, THF,† $-78 \rightarrow -60$ °C, then aq. NH₄Cl, 75%) afforded the diacetylene (9)† (mixture of isomers), deprotection of which (AgNO₃–KCN, EtOH–H₂O, 0 °C, 94%) led to compound (10). Sequential alkylation of (10) {a, 1.0 equiv. of BuⁿLi, THF, Me[CH₂]₄I, HMPA,† $-40 \rightarrow -25$ °C, 82%; b,⁵ 1.0 equiv. of BuⁿLi, THF, (MeO)₃C[CH₂]₃I, HMPA, $-40 \rightarrow -25$ °C, mild acid hydrolysis, and flash silica column chromatography, 75% yield based on *ca*. 75% conversion} furnished the desired precursor (12).

Thermolysis³ of (12) (200 °C, benzene, sealed tube) smoothly gave rise to the two products (4; R=Me) and (5; R=Me) in essentially quantitative yield [racemic, (4):(5) ca. 4:1], separated by flash silica column chromatography [(4; R=Me):† R_f 0.18, 3% ether in light petroleum, four developments; (5; R=Me):‡ R_f 0.20, 3% ether in light petroleum, four developments].

‡ All new compounds gave satisfactory spectroscopic and analytical data: ¹H n.m.r., CDCl₃, 250 MHz: (2; R=Me) δ 0.88 (3H, t, J 6.7 Hz 20-H), 1.19—1.42 (6H, m, 17-, 18-, and 19-H), 1.70 (2H, m, 3-H), 1.95—2.18 (4H, m, 4- and 16-H), 2.32 (2H, t, *J* 7.5 Hz, 2-H), 2.66 (1H, dt, J 17.5 and 7.0 Hz, 10-H), 3.12 (1H, br. d, J 17.5 Hz, 10-H), 3.47 (2H, m, 7- and 13-H), 3.66 (3H, s, CO₂Me), and 5.30—5.72 (8H, m, olefinic); (3; R=Me) δ 0.86 (3H, t, J 6.7 Hz, 20-H), 1.23—1.39 (6H, m, 17-, 18-, and 19-H), 1.66 (2H, m, 3-H), 1.96—2.12 (4H, m, 4- and 16-H), 2.29 (2H, t, J 7.5 Hz, 2-H), 2.85 (2H, m, 10-H), 3.46 (2H, m, 7and 13-H), 3.64 (3H, s, CO₂Me), and 5.20—5.70 (8H, m, olefinic); (4; R=Me), δ 0.87 (3H, t, J 6.7 Hz, 20-H), 1.21—1.56 (6H, m, 17-, 18-, and 19-H), 1.79 (2H, m, 3-H), 2.16 (2H, dt, J7.5 and 2.9 Hz, 16-H), 2.24 (2H, dt, J 7.0 and 2.0 Hz, 4-H), 2.46 (2H, t, J 7.5 Hz, 2-H), 2.10 and 2.93 (each 1H, br. d, J 17.5 Hz, 10-H), 3.54 (2H, br. s, 7- and 13-H), 3.64 (3H, s, CO₂Me), and 5.66 (4H, m, 8-, 9-, 11-, and 12-H); (5; R=Me) δ 0.88 (3H, t, J 6.7 Hz, 20-H), 1.18—1.56 (6H, m, 17-, 18-, and 19-H), 1.8 (2H, m, 3-H), 2.17 (2H, t, J 7.5 Hz, 16-H), 2.25 (2H, t, J 7.0 Hz, 4-H), 2.84 (2H, br. s, 10-H), 3.45 (2H, br. s, 7- and 13-H), 3.65 (3H, s, CO₂Me), and 5.68 (4H, m, 8-, 9-, 11-, and 12-H).

[†] Abbreviations: THF = tetrahydrofuran, HMPA = hexamethylphosphoramide.

The stereochemistry of the 7,13 junction in (4) and (5) was based on the $^1\mathrm{H}$ n.m.r. signals for H-10 according to the assignments of Baldwin *et al.*³ Controlled hydrogenation of the *cis*-isomer (4; R=Me) (Lindlar catalyst, hexane, 25 °C) resulted in the formation of (2; R=Me)‡ (70%; R_{f} 0.35, 10% ether in light petroleum), whereas similar treatment of the *trans*-isomer (5) furnished (3; R=Me)‡ (70%; R_{f} 0.38, 10% ether in light petroleum, silica). Finally mild alkaline hydrolysis of (2)—(4) (R=Me) (LiOH-H₂O-THF, 25 °C) led to the arachidonic acids (2)—(4) (R=H) in almost quantitative yield.

Preliminary biological and chemical investigations with these newly synthesized molecules indicate high stability and potent and selective 5-lipoxygenase inhibitory activity.

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References

- Selected reviews: B. Samuelsson, M. Goldyne, E. Granstrom, M. Hamberg, S. Hammarstrom, and C. Malmsten, Ann. Rev. Biochem., 1978, 47, 997; K. C. Nicolaou and J. B. Smith, Ann. Rep. Med. Chem., 1979, 14, 178; T. K. Shaaf, ibid., 1977, 12, 182; E. J. Corey, Experientia, 1982, 38, 1259; B. Samuelsson, Pure Appl. Chem., 1981, 53, 1203; Angew. Chem., Int. Ed. Engl., 1982, 21, 902; D. M. Bailey and F. B. Casey, Ann. Rep. Med. Chem., 1982, 17, 203; D. M. Bailey and L. W. Chakrin, ibid., 1981, 16, 2; P. Borgeat and P. Sirois, J. Med. Chem., 1981, 24, 121.
- E. J. Corey, H. Niwa, and J. R. Falck, J. Am. Chem. Soc., 1981, 101, 1586.
- 3 J. E. Baldwin and C. Ullenius, J. Am. Chem. Soc., 1974, 96, 1546, and references cited therein.
- 4 E. R. H. Jones, M. Ahmed, G. Barley, M. T. W. Hearn, V. Thaller, and J. Yates, J. Chem. Soc., Perkin Trans. 1, 1974, 1981.
- 5 E. J. Corey and J. Kang, Tetrahedron Lett., 1982, 23, 1654.